

## *pEASY*<sup>®</sup>-T1 Cloning Kit

Please read the user manual carefully before use.

Cat. No. CT101

### Storage

*Trans*1-T1 Phage Resistant Chemically Competent Cell at -70°C for six months; others at -20°C for nine months

### Descriptions

*pEASY*<sup>®</sup>-T1 Cloning Kit is designed for cloning and sequencing *Taq*-amplified PCR products.

- 5 minutes fast ligation of *Taq*-amplified PCR products.
- Kanamycin and Ampicillin resistance genes for selection.
- Easy blue/white selection.
- T7 promoter, M13 forward and M13 reverse primers for sequencing.
- T7 promoter for *in vitro* transcription.
- *Trans*1-T1 Phage Resistant Chemically Competent Cells, high transformation efficiency (>10<sup>9</sup> cfu/μg pUC19 DNA) and fast growing.

### Kit Contents

Component	CT101-01 (20 rxns)	CT101-02 (60 rxns)
<i>pEASY</i> <sup>®</sup> -T1 Cloning Vector (10 ng/μl)	20 μl	3×20 μl
Control Template (5 ng/μl)	5 μl	5 μl
Control Primers (10 μM)	5 μl	5 μl
M13 Forward Primer (10 μM)	50 μl	150 μl
M13 Reverse Primer (10 μM)	50 μl	150 μl
<i>Trans</i> 1-T1 Phage Resistant Chemically Competent Cells	10×100 μl	30×100 μl

### Preparation of PCR Products

1. Primer requirement: primer cannot be phosphorylated
2. PCR Enzyme: *Taq* DNA polymerases
3. Reaction conditions: in order to ensure the integrity of amplification products, 5-10 minutes of post-extension step is required.  
After amplification reaction, use agarose gel electrophoresis to verify the quality and quantity of PCR product

### Setting Up the Cloning Reaction System

Add following components into a microcentrifuge tube.

PCR products                                    0.5-4 μl (can be increased or reduced based on PCR product yield, no more than 4 μl)

*pEASY*<sup>®</sup>- T1 Cloning Vector                1 μl

Gently mix well, incubate at room temperature (20°C-37°C) for 5 minutes. After reaction, place the tube on ice.

#### 1. Optimal amount of insert

Molar ratio of vector to insert = 1:7 (1 kb, ~20 ng; 2 kb, ~40 ng)

#### 2. Optimal volume of vector: 1 μl

#### 3. Optimal reaction volume: 3~5 μl

#### 4. Optimal incubation time

(1) 0.1~1 kb (including 1 kb): 5~10 minutes

(2) 1~2 kb (including 2 kb): 10~15 minutes

(3) 2~3 kb (including 3 kb): 15~20 minutes

(4) ≥3 kb: 20~30 minutes

Use the maximum incubation time if the insert is gel purified.

- Optimal incubation temperature: for most PCR inserts, the optimal temperature is about 25°C; for some PCR inserts, optimal results can be achieved with higher temperature (up to 37°C).

#### Transformation

- Add the ligated products to 50 µl of *Trans1-T1* Phage Resistant Chemically Competent Cell and mix gently (do not mix by pipetting up and down).
- Incubate on ice for 20~30 minutes.
- Heat-shock the cells at 42°C for 30 seconds.
- Immediately place the tube on ice for 2 minutes.
- Add 250 µl of room temperature SOC or LB medium. Shake the tube at 37°C (200 rpm) for 1 hour.
- In the meantime, mix 8 µl of 500 mM IPTG with 40 µl of 20 mg/ml X-gal. Spread them evenly onto a selective LB plate. Place the plate at 37°C for 30 minutes.
- Spread 200 µl or all transformants on the pre-warmed plate. Incubate at 37°C overnight.

#### Identification of Positive Clones and Sequencing

##### Analysis of positive clones

- Transfer 5~10 white or light blue colonies into 10 µl ddH<sub>2</sub>O and vortex.
- Use 1 µl of the mixture as template for 25 µl PCR using M13 forward and M13 reverse primers.
- PCR reaction conditions

94°C	10 min	} 30 cycles
94°C	30 sec	
55°C	30 sec	
72°C	x min*	
72°C	5-10 min	

\* (depends on the insert size and PCR enzymes) the PCR product size from vector self-ligation is 199 bp.

- Analyze positive clones by restriction enzyme digestion and DNA sequencing.  
Inoculate positive clones on LB/Amp<sup>+</sup> or LB/Kan<sup>+</sup> liquid medium, grow at 37°C for 6 hours at 200 rpm. Isolate plasmid DNA by plasmid MiniPrep Kit. Analyze colonies by restriction enzyme digestion with proper restriction endonuclease.

#### Sequencing

Analyze the sequence by sequencing with M13 F, M13 R and T7 promoter.

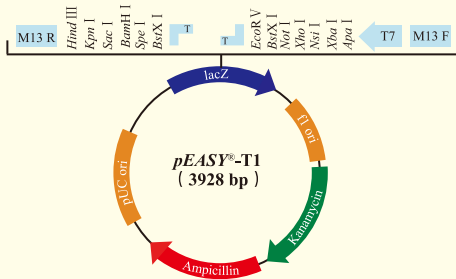
#### PCR for control insert (700 bp)

Component	Volume	Final Concentration
Control Template (5 ng/µl)	1 µl	0.1 ng/µl
Control Primers (10 µM)	1 µl	0.2 µM
2× <i>EasyTaq</i> <sup>®</sup> PCR SuperMix	25 µl	1×
ddH <sub>2</sub> O	Variable	-
Total volume	50 µl	-

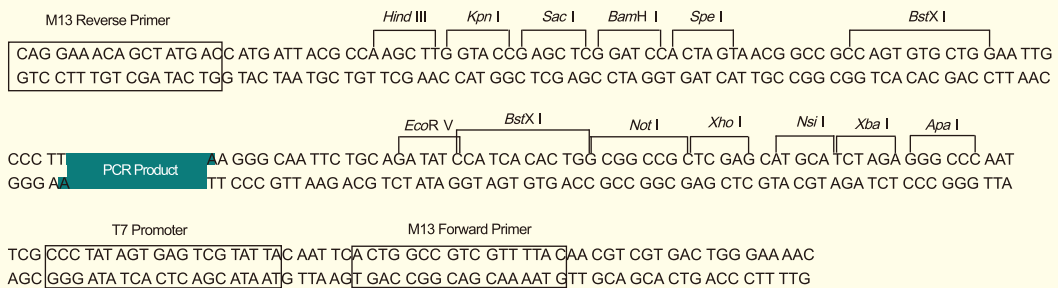
#### Thermal cycling conditions for control insert

94°C	2-5 min	} 30 cycles
94°C	30 sec	
55°C	30 sec	
72°C	1 min	
72°C	10 min	

Ligate 1 µl of control PCR insert with 1 µl vector. Hundreds of colonies should be produced with cloning efficiency over 90%.



*LacZa* fragment: bases 1-544  
 M13 reverse priming site: bases 205-221  
 Multiple cloning site: bases 234-354  
 T7 promoter priming site: bases 361-380  
 M13 forward priming site: bases 387-403  
 f1 origin: bases 545-982  
 Kanamycin resistance ORF: bases 1,316-2,110  
 Ampicillin resistance ORF: bases 2,128-2,988  
 pUC origin: bases 3,133-3,806



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