



### FavorPrep™ Tissue Total RNA Mini Kit

-- For isolation RNA from animal cells, animal tissues, bacteria, yeast, paraffin fixed sample, fungi and for RNA clean-up

Cat. No.: FATRK 000  
FATRK 001  
FATRK 001-1  
FATRK 001-2  
For Research Use Only

#### Kit Contents:

Cat. No:	FATRK 000-Mini (4 preps_sample)	FATRK 001 (50 preps)	FATRK 001-1 (100 preps)	FATRK 001-2 (300 preps)
FARB Buffer	3 ml	25 ml	45 ml	130 ml
Wash Buffer 1	3 ml	30 ml	60 ml	170 ml
Wash Buffer 2 (concentrate) <sup>a</sup>	1.5 ml	15 ml	35 ml	50 ml x 2
RNase-free Water	0.5 ml	6 ml	6 ml	8 ml x 2
Filter Column	4 pcs	50 pcs	100 pcs	300 pcs
FARB Mini Column	4 pcs	50 pcs	100 pcs	300 pcs
Collection Tube	8 pcs	100 pcs	200 pcs	600 pcs
Elution Tube	4 pcs	50 pcs	100 pcs	300 pcs
Micropestle	4 pcs	50 pcs	100 pcs	300 pcs
User Manual	1	1	1	1
Preparation of Wash Buffer by adding ethanol (96 ~ 100%)				
Ethanol volume for Wash Buffer 2 <sup>a</sup>	6 ml	60 ml	140 ml	200 ml

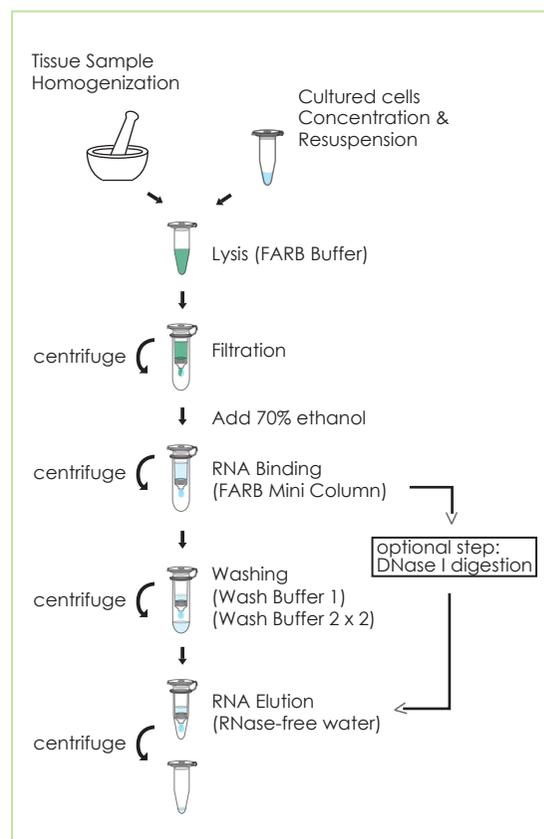
#### Specification:

Principle: mini spin column (silica matrix)  
Operation time: 30 ~ 60 minutes  
Binding capacity: up to 100 µg total RNA/ column  
Column applicability: centrifugation and vacuum  
Minimum elution volume: 40 µl

#### Sample amount and yield

Sample	Recommended amount of sample used	Yield (µg)
Animal cells (up to 5 x 10 <sup>6</sup> )	NIH/3T3	10
	HeLa	15
	COS-7	30
	LMH	12
Animal Tissue (Mouse/rat) (up to 30 mg)	Embryo	25
	Heart	10
	Brain	10
	Kidney	30
	Liver	50
	Spleen	35
	Lung	15
Thymus	45	
Bacteria	E. coli	60
	B. subtilis	40
Yeast (up to 5 x 10 <sup>7</sup> )	S. cerevisiae	25

#### Brief procedure:



#### Important Notes:

1. Make sure everything is RNase-free when handling RNA.
2. Buffers provided in this system contain irritants. Wear gloves and lab coat when handling these buffers.
3. **Caution: β-mercaptoethanol (β-Me) is hazardous to human health. perform the procedures involving β-Me in a chemical fume hood.**
4. Add required volume of RNase-free ethanol (96~100%) to Wash Buffer 2 when first use.
5. All centrifuge steps are done at full speed (~18,000 x g) in a microcentrifuge.
6. Prepare RNase-free DNase 1 reaction buffer (1M NaCl, 10 mM MnCl<sub>2</sub>, 20 mM Tris-HCl, pH 7.0 at 25°C) and make the final concentration of DNase I to 0.5 U/µl.

#### Protocol: Isolation of Total RNA from Animal Cells

Please Read Important Notes Before Starting Following Steps.

Additional requirement: β-Mercaptoethanol  
70% RNase-free ethanol

1. Collect 1 ~ 5 × 10<sup>6</sup> cells by centrifuge at 300 x g for 5 min at 4 °C. Remove all the supernatant.  
**-- Note! Do not overload, too much sample will make cell lysis incompletely and lead to lower RNA yield and purity.**
2. Add 350 µl of FARB Buffer and 3.5 µl of β-Mercaptoethanol to the cell pellet. Vortex vigorously for 1 min to resuspend the cells completely.  
**-- Note: If the clump is still visible after vortex, pipet sample mixture up and down to break down the clump.**
3. Place a Filter Column to a Collection Tube and transfer the sample mixture to the Filter Column. Centrifuge at full speed (~ 18,000 x g) for 2 min.
4. Transfer the clarified supernatant from the Collection Tube to a new microcentrifuge tube (not provided), and measure the volume of the supernatant.  
**-- Note: Avoid to pipet any debris and pellet when transferring the supernatant.**
5. Add 1 volume of 70 % RNase-free ethanol and mix well by vortexing.
6. Place a FARB Mini Column to a Collection Tube and transfer the ethanol added sample mixture (including any precipitate) to the FARB Mini Column. Centrifuge at full speed for 1 min, discard the flow-through and return the FARB Mini Column back to the Collection Tube.
7. **Optional step: DNase I digestion** To eliminate genomic DNA contamination, follow the steps from 7a.  
Otherwise, proceed to step 8 directly.
  - 7a. Add 250 µl of Wash Buffer 1 to the FARB Mini Column, centrifuge at full speed for 1 min. Discard the flow-through and return the FARB Mini Column back to the Collection Tube.
  - 7b. Add 60 µl of RNase-free DNase 1 solution (0.5U/ul, not provided) to the membrane center of the FARB Mini Column. Place the column on the benchtop for 15 min.
  - 7c. Add 250 µl of Wash Buffer 1 to the FARB Mini Column, centrifuge at full speed for 1 min. Discard the flow-through and return the FARB Mini Column back to the Collection Tube.
  - 7d. After DNase 1 treatment, proceed to step 9.
8. Add 500 µl of Wash Buffer 1 to the FARB Mini Column, centrifuge at full speed for 1 min. Discard the flow-through and return the FARB Mini Column back to the Collection Tube.
9. Add 750 µl of Wash Buffer 2 to the FARB Mini Column, centrifuge at full speed for 1 min. Discard the flow-through and return the FARB Mini Column back to the Collection Tube.  
**-- Note: Make sure that ethanol has been added into Wash Buffer 2 when first use.**
10. Repeat step 9 for one more washing.
11. Centrifuge the FARB Mini Column at full speed for an additional 3 min to dry the FARB Mini Column.  
**-- Important Step! This step will avoid the residual liquid to inhibit subsequent enzymatic reaction.**
12. Place the FARB Mini Column to a Elution Tube (provided, 1.5 ml microcentrifuge tube).
13. Add 40 ~ 100 µl of RNase-free ddH<sub>2</sub>O to the membrane center of the FARB Mini Column. Stand the FARB Mini Column for 1 min.  
**-- Important Step! For effective elution, make sure that RNase-free ddH<sub>2</sub>O is dispensed on the membrane center and is absorbed completely.**  
**-- Important : Do not elute the RNA using RNase-free water less than suggested volume (< 40 µl). It will lower the RNA yield.**
14. Centrifuge the FARB Mini Column at full speed for 1 min to elute RNA.
15. Store RNA at -70C.

### Protocol: Isolation of Total RNA from Animal Tissues

#### Please Read Important Notes Before Starting Following Steps.

Additional equipment: liquid nitrogen & mortar  
a rotor-stator homogenizer or a 20-G needle syringe  
β-Mercaptoethanol  
70% RNase-free ethanol

A-1 Weight up to 30 mg of tissue sample. Grind the sample in liquid nitrogen to a fine powder with a mortar and transfer the powder to a new microcentrifuge tube (not provided).

-- **Note! Avoid thawing the sample during weighing and grinding.**

A-2. Add 350 μl of FARB Buffer and 3.5 μl of β-Mercaptoethanol. Homogenize the sample by using a rotor-stator homogenizer or by passing the sample lysate through a 20-G needle syringe 10 times. Incubate at room temperature for 5 min.

-- **Important step: In order to release more RNA from the harder samples, it is recommended to homogenize the sample by using suitable homogenize equipment, for example, with a rotot-stator homogenizer.**

A-3. Follow the Animal Cells Protocol starting from step 3.

#### **(Alternative)**

B-1. Place up to 30 mg of tissue sample to a microcentrifuge tube. Add 350 μl of FARB Buffer and 3.5 μl of β-Mercaptoethanol and use provided micropestle to grind the tissue sample thoroughly.

B-2. Homogenize the sample by passing lysate through a 20-G needle syringe 10 ~ 20 times. Incubate at room temperature for 5 min.

-- **For the tissue samples having low cell amount and hard to disrupt, it is recommended to proceed A1-A3 step above.**

B-3. Follow Animal Cells Protocol starting from step 3.

### Protocol: Isolation of Total RNA from Bacteria

#### Please Read Important Notes Before Starting Following Steps.

Additional requirement: β-Mercaptoethanol  
70% RNase-free ethanol  
30 °C water bath or heating block  
2 ml screw centrifuge tube  
Lysozyme reaction solution: (10mg/ ml lysozyme; 20mM Tris-HCl, pH 8.0; 2mM EDTA;  
1.2% Triton)  
Acid-washed glass beads, 500 ~ 700 μm

1. Transfer up to  $1 \times 10^9$  cells well-grown bacterial culture to a 2 ml screw centrifuge tube.

-- **Note! Make sure the amount of total RNA harvested from sample do not exceed the column's binding capacity (100 μg) when estimate the sample size. Too much sample will make cell lysis incompletely and lead to lower RNA yield and purity. If RNA amount is hard to determine on some species, using  $\leq 5 \times 10^8$  cells as the starting sample size.**

2. Descend the bacterial cells by centrifuge at full speed (~18,000 x g) for 2 min at 4 °C. Remove all the supernatant.

3. Add 100 μl of lysozyme reaction solution. Pipet up and down to resuspend the cell pellet and incubate at 37°C for 10 min.

4. Add 350 μl of FARB Buffer and 3.5 μl of β-Mercaptoethanol.

5. Add 250 mg of acid-washed glass beads (500 ~ 700 nm) and vortex vigorously for 5 min to disrupt the cells.

6. Centrifuge at full speed (~18,000 x g) for 2 min to spin down insoluble material. Transfer the supernatant to a microcentrifuge tube (not provided) and measure the volume of the clear lysate.

-- **Note! Avoid pipetting any debris and pellet in the Collection Tube.**

7. Follow Animal Cells Protocol starting from step 5.

### Protocol: Isolation of Total RNA from Yeast

#### Please Read Important Notes Before Starting Following Steps.

Additional requirement: β-Mercaptoethanol  
70% RNase-free ethanol

Enzymatic disruption: Lyticase or zymolase  
Sorbitol buffer (1 M sorbitol; 100 mM EDTA; 0.1% β-ME )  
30 °C water bath or heating block

Mechanical disruption: 2 ml screw centrifuge tube  
Acid-washed glass beads, 500 ~ 700 μm

1. Collect up to  $5 \times 10^7$  of yeast culture by centrifuge at 5,000 x g for 10 min at 4 °C. Remove all the supernatant.

2A. Enzymic disruption:

2A-1: Resuspend the cell pellet in 600 μl sorbitol buffer (1 M sorbitol; 100 mM EDTA; 0.1% β-ME) (not provided). Add 200 U zymolase or lyticase and incubate at 30 °C for 30 min.

--**Note! Prepare sorbitol buffer just before use.**

2A-2. Centrifuge at 300 x g for 5 min to pellet the spheroplasts. Remove all the supernatant.

2A-3. Add 350 μl of FARB Buffer and 3.5 μl of β-Mercaptoethanol to the pellet. Vortex vigorously to disrupt the spheroplasts for 1 min. Incubate sample mixture at room temperature for 5 min.

2B. Mechanical disruption:

2B-1. Add 350 μl of FARB Buffer and 3.5 μl of β-Mercaptoethanol to the pellet and vortex vigorously to resuspend the cells completely.

2B-2. Transfer the sample mixture to a 2 ml screw centrifuge tube and add 250 mg of acid-washed glass beads (500 ~ 700 nm) and vortex vigorously for 15 min to disrupt the cells.

3. Follow Animal Cells Protocol starting from step 5.

### Protocol: Isolation of Total RNA from Paraffin-embedded tissue

#### Please Read Important Notes Before Starting Following Steps.

Additional equipment: xylene & ethanol (96~100%)  
liquid nitrogen & mortar  
a rotor-stator homogenizer or a 20-G needle syringe  
β-Mercaptoethanol  
70% RNase-free ethanol

1. Transfer up to 15 mg paraffin-embedded tissue sample to a microcentrifuge tube (not provided).

-- Remove the extra paraffin to minimize the size of the sample slice.

2. Add 0.5 ml xylene, mix well and incubate at room temperature for 10 min.

3. Centrifuge at full speed for 3 min. Remove the supernatant by pipetting.

4. Add 0.25 ml xylene, mix well and incubate at room temperature for 3 min.

5. Centrifuge at full speed for 3 min. Remove the supernatant by pipetting.

6. Repeat step 4 and step 5

7. Add 0.3 ml ethanol (96- 100 %) to the deparaffined tissue, mix gently by vortexing. Incubate at room temperature for 3 min.

8. Centrifuge at full speed for 3 min. Remove the supernatant by pipetting.

9. Repeat step 7 and step 8.

10. Follow Animal tissue Protocol starting from step 1 for sample disruption then follow Animal Cells protocol starting from step 3.

### Protocol: RNA clean up

#### Please Read Important Notes Before Starting Following Steps.

Additional equipment: xylene & ethanol (96~100%)

1. Transfer 100 μl of RNA sample to a microcentrifuge tube (not provided).

-- If the RNA sample is less than 100 μl, add RNase-free water to make the sample volume to 100 μl.

2. Add 300 μl of FARB Buffer and 300 μl of RNase-free ethanol (96~100 %) and mix well by vortexing.

3. Place a FARB Mini Column to a Collection Tube and transfer the ethanol added sample mixture to the FARB Mini Column. Centrifuge at full speed for 1 min and discard the flow-through and return the FARB Mini Column back to the Collection Tube.

4. Follow Animal Cells Protocol starting from step 8.